Cytotoxic Effect of a Mycelial extracts on Different Cell lines and Experimental animals

Osman, M.E.¹, Wafaa Ahmed², Heba, E.E.¹ and Om Hashim. M³

Abstract-Mushroom polysaccharides play an important role as functional foods because they exhibit biological modulator properties such as antitumor, antiviral and antibacterial activities. Cytotoxic and antitumor activities of hot aqueous extracts of five macrofungi were studied. Hot water extracts showed in vitro cytotoxic activity against Breast carcinoma cell line (MCF7), Liver carcinoma cell line (Hep-G2) and Colon carcinoma cell line (HCT116). Flammulina velutipes 6 polysaccharide extract had potential for tumor therapy. So, it was selected for in vivo studies. The antitumor activity of F. velutipes 6 polysaccharide extract against Ehrlich carcinoma tumor in the mice model was significant. Significant regression of tumor growth was observed in the mice upon oral administration of 200 mg/kg F. velutipes 6 polysaccharide extract for 10 days before tumor implantation with inhibition ratio of 99.34% (mean tumor volume was 0.006mm³), while 4 out of 15 mice showed complete tumor regression. While, group with oral injection after tumor implantation of F. velutipes 6 polysaccharide extract inhibited tumor growth was even more effective than group injected with extract before tumor implantation. The inhibition ratio of 99.79 % (mean tumor volume was 0.002mm³), while 5 out of 15 mice showed complete tumor regression in this group.

Keywords—Cytotoxicity, Experimental animals, Mushroom, Polysaccharide, Tumor induction.

I. INTRODUCTION

Mushrooms have been consumed globally as tasty food and nutritional supplements. The medicinal power and nutritional value of several mushrooms are widely known. However, only since the last decade of the 20th century, it has been possible to isolate and partially characterize some biologically active antitumor substances [1].

The therapeutic properties of mushrooms are attributed mainly to their content of polysaccharides, which exhibit biological modulator properties such as antitumor, antiviral and antibacterial activities [2]–[6].

From a structural view point, polysaccharides are a diverse group of biological macromolecules that are found to occur in several organisms. They contain repetitive structural features and are polymers of monosaccharide residues joined to each other by glycosidic bonds. Therefore they differ structurally from proteins and nucleic acids and have the highest capacity for carrying biological information since they have the greatest potential for structural variability [1],[2].

The effects of mushroom nutrition on clinical conditions have attracted great interest in the scientific community in the last decade in order to understand the molecular mechanism responsible for their action [7].

Protein-bound polysaccharides are macromolecules that consist of a central core protein to which are attached a number of polysaccharide chains. Polysaccharide peptide (PSP) and polysaccharide-K (PSK) from Coriolus versicolor are among the most widely studied protein-bound polysaccharides. However, such basidiomycetes also synthesize free polysaccharides that exhibit powerful biological activity [8, 9].

There are some reports on the production of free and protein-bound polysaccharides from basidiomycete strains such as C. versicolor, Pleurotus ostreatus, Lentinula edodes and Ganoderma lucidum [2], [10], [11].

Flammulina velutipes (F.velutipes) is a cultivated mushroom. Few studies, however, have been conducted on this species. An alkaline protease and the antitumor activities have been reported from this mushroom [12], [13]. Both methanolic and ethyl acetate extracts of this mushroom exhibited anti-hyperlipidemic and antioxidant activities [14]. As result of its perceived health benefits, F. velutipes has become one of the valuable mushrooms in China.

Therefore the aim of this study was a trial to produce polysaccharides from different mushroom species grown by submerged culture and studying their cytotoxic effect on human cancer cell lines" in vitro study". Then the most potent extract was studied on animal model "in vivo"

II. MATERIALS AND METHODS

Fungal Strains:

Five mushroom strains Lentinus edodes LC2141, Lentinus edodes LC202, Flammulina velutipes 5, Flammulina velutipes 6 and Flammulina velutipes 13 were kindly obtained from Fujian Agriculture Univ., China. All strains were maintained on Potato Dextrose Agar (PDA) slants and incubated at 25°C for 7 days then stored in refrigerator at 4°C after growth for routine culture and storage purposes.

Culture Media and culture conditions:

All fungal strains were initially grown on PDA medium in Petri dishes for 7–8 days. The flask culture experiments were performed in a 250 ml flask containing 50 ml of mushroom fermentation medium (MFM). Each flask was inoculated with

¹Department of Botany and Microbiology, Faculty of Science, Helwan University, Egypt.

²Department of Cancer Biology, National Cancer Institute, Cairo University, Egypt.

³Biomedical Engineering Department- High Institute of Engineering, el-Shorouk-Academy Egypt.

10 agar plugs 5 mm2 of the agar plate culture with sterilized cork porer. MFM medium consisted of the following components (g/liter): glucose (35), peptone (5), yeast extract (5), KH2PO4·H2O (1), MgSO4·7H2O (0.5) and vitamin B1 (0.05) [15].

Determination of dry weight:

The fungal mats were separated by filtration and washed several times with distilled water then dried at 70°C to a constant weight.

Preparation of crude hot water extract:

The mycelia of the five tested strains were dried at 60° C before analysis. The dried mycelia were pulverized and 5.0g of the powdered sample were extracted with 80% (v/v) ethanol for 24 h, and then filtered. The residues were dried and extracted with distilled water at 100 °C three times. The whole extract was filtered and centrifuged. After centrifugation (6000 rpm for 10min), The supernatant was concentrated by evaporation under reduced pressure and treated with three volumes of ethanol for precipitation at 4 °C overnight. The precipitate was obtained by centrifugation, and dried to give a crude extract [16].

Human tumor cell lines and in vitro cytotoxicity assay:

Human cancer cell lines which used in this study, were MCF7 (Breast carcinoma cell line), Hep-G2 (Liver carcinoma cell line) and HCT116 (Colon carcinoma cell line). They were obtained frozen in liquid nitrogen (-180oC) from the American Type Culture Collection. The tumor cell lines were maintained in the National Cancer Institute, Cairo, Egypt, by serial sub-culturing. Cytotoxicity was determined using the MTT assay. For the assessment of the cytotoxic and cytostatic activities of all mushroom extracts, cells were seeded in 96well flat-bottomed microtiter plates at a density of approximately (0.5X105 cells/well), in complete RPMI-1640 Medium. After 24 h to ensure cell attachment, serial dilutions of the extracts in physiological saline were prepared. 100 µl of different concentrations of each tested extracts were added for 24 h at 37°C, in a humidified 5 % CO2 atmosphere. After incubation, medium was removed, wells washed with 100ul PBS then 30 µl MTT solution/well was added and incubated for an additional 4 h. MTT crystals were solubilized by added 50µl of DMSO/well then the plate was shacked at room temperature. It was followed by photometric determination of the absorbance at 570 nm using microplate ELISA reader after development of violet color. Control cells were treated with vehicle alone. For each compound concentration, 3 wells were used (triplicate wells were prepared for each individual dose). The average was calculated. The half maximal inhibitory concentration (IC50) was determined after plotting the dose response curve for each conc. using Graph pad Prism software. The inhibition rate was calculated according to the formula below:

Inhibition rate (%) =1- (Abs _{sample}/Abs _{control}) × 100

Antitumor activity "In vivo" experiments:

• Experimental Animals

The experimental animals used in this study were adult male albino mice weighing from 19-20 g purchased from the animal house colony of the National Cancer Institute (NCI), Cairo University, Egypt. Animals were housed under normal environmental conditions of standard temperature, humidity and diurnal environment of light and dark in suitable cages and kept on standard diet.

• Experiment design:

Animals were divided into five groups:

Group (1): Ten mice were considered as a negative control group which was not inoculated with Ehrlich tumor cells. Control mice were orally administered with physiological saline only.

Group (2): 15 mice were injected with Ehrlich carcinoma cells only (positive control).

Group (3):15 mice-bearing Ehrlich carcinoma cells were orally administered with 0.2 ml of the hot water extract from mycelia of strain Flammulina velutipes 6 each for 10 days at a daily dose 200 mg/kg started after 24h of tumor implantation. "Treatment"

Group (4):15 mice were orally administered with 0.2 ml of the hot water extract from mycelia of strain Flammulina velutipes 6 each for 10 days at a daily dose 200 mg/kg and then implanted with Ehrlich carcinoma cells. "Protective"

Group (5):10 mice were orally administered with 0.2 ml of the hot water extract from mycelia of strain Flammulina velutipes 6 each for 10 days at a daily dose 200 mg/kg only.

• Solid tumor production:

Solid tumors were produced by intramuscular inoculation in the right thigh of the lower limb of each mouse with 0.1 ml of Ehrlich carcinoma cells, which contained 1 x 106 viable Ehrlich carcinoma cells. Viability, assessed by the Trypan blue dye exclusion method, was found to be 95% or more.

• Evaluation of the antitumor activity:

Antitumor activity was assessed by measuring the change in tumor volume (TV/mm3), body weight (BW/g) and survival rate.

Tumor volume (Size) (TV) measurements:

The change in tumor volume (TV/mm3) was measured at different time intervals during the experiment time course using a Vernier caliper and calculated by the following equation:. The tumor volume was calculated using the formula:

Tumor volume (mm3) = $4/3\pi a2b/2$, Where a is the smallest and b is the largest diameter (mm). $\pi = 22/7$

Body weight changes:

Body weight (BW/g) and survival were monitored throughout the experimental time course.

Antitumor inhibition ratio (%) [17]:

=100 (1-T/C)

Where

C is the average tumor size (mm3) in control group. T is the average tumor size (mm3) in Treatment group.

Statistical Analysis:

Statistical analysis of data was carried out by using one way analysis of variance (ANOVA) followed by homogenous subsets (Duncan) at confidence level of 5% (0.05) using the Statistical Package for the Social Science (SPSS) version17. Duncan's multiple range tests were used to compare between means of treatments.

III. RESULTS AND DISCUSSION

In vitro cytotoxicity assay:

The treatment options of cancer including surgery, chemotherapy, radiation therapy, and palliative care often are expensive and have side effects. For example, most chemotherapeutic agents for the treatment of cancer destroy tumors and stop cancer progress but also damage healthy cells and tissues. Due to these side effects, researchers search for new antitumor substances from various natural sources, especially fungi [18]-[20]. More than 140 fungal metabolites have shown confirmed activity in tumor cell line bioassays [21].

Many kinds of polysaccharides have shown significant antitumor activities and low side-effects in vivo. Most reports conformed that mushroom polysaccharides exerted their antitumor action via activation of the immune response of the host organism, and mushroom polysaccharides were regarded as biological response modifiers (BRMs). However, in the past few years, studies related with the effects of polysaccharides on the tumor cells have increased. It has also been reported that polysaccharides have different antitumor activities in vitro, depending on their monosaccharide composition, protein contents, molecular mass, and chain conformation [22].

The antitumor activity of the tested polysaccharide extracts against the three carcinoma cell lines is shown in (table 1A-E). Cytotoxicity was measured and expressed as the survival fraction compared with untreated control cells, where the cell viability was determined, and IC50 was determined from a

range of concentrations shown in Figures 1. According to the in vitro inhibition ratio of MCF7 breast cell line by the five polysaccharide extract of mushrooms mycelia (Lentinus edodes LC2141, Lentinus edodes LC202, Flammulina velutipes 5, Flammulina velutipes 6 and Flammulina velutipes 13) at different concentrations (2.5, 5, 10 and 20 μ g/ml) (fig.1), all extracts exhibited strong inhibition against cell growth at all concentrations, and there was also a dose–response relationship between concentration of the mycelial polysaccharides and suppression of MCF7 cell proliferation. The polysaccharide extract of Lentinus edodes LC2141 mycelia showed the highest significant antitumor effect as compared with other polysaccharide extracts from the tested mushroom strains. It was noted that extract of Lentinus edodes LC2141 showed high inhibition ratio of 72.3 % at the concentration of 10µg/ml. While the extracts of Lentinus edodes LC202 and Flammulina velutipes 6 showed inhibition ratio of 60.77 % and 53.3% at the concentration of 20µg/ml. The extracts of both Flammulina velutipes 5 and Flammulina velutipes 13 exhibited no cytotoxic effect with all tested concentrations. The treatment of MCF7 cells with different concentrations of hot water extract of L. edodes LC2141, L. edodes LC202 and F. velutipes 6 dramatically inhibited the cell growth with IC50 values of 7.5µg/ml, 4.5µg/ml and 5µg/ml respectively.

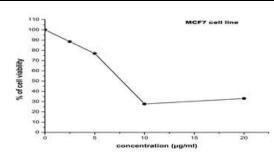
Also, the treatment of Hep-G2 liver cell line with the mushroom polysaccharide extracts at different concentrations revealed that all extracts exhibited strong inhibitory effect against cell growth at all concentrations (fig.2). The extracts of L. edodes LC202, F. velutipes 6, F. velutipes 13, F. velutipes 5 and L. edodes LC2141 exhibited high inhibition ratio of 67.88%, 67.60%, 66.20%, 55.03% and 53.35% respectively at the concentration of 2.5µg/ml for L. edodes LC202 extract and 20µg/ml for the other strains. The polysaccharide extracts of L. edodes LC2141 and L. edodes LC202 inhibited the cell growth with IC50 values of 5µg/ml. *F.* velutipes 6 and F. velutipes5 showed cell inhibition with IC50 values of 4µg/ml and 8µg/ml respectively.

Table 1: Inhibition ratios of different cancer cell lines with IPS extract of *the five studied Mashrom strain using* concentration range from 2.5 to 20µg/ml(A-E)

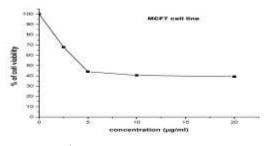
		А				
	Inhibition ratio %	with IPS extract	of Lentinula			
CONC.	edodes LC2141					
µg/ml	MCF7	Hep-G2	HCT116			
		-				
2.5	11.54	26.82	14.01			
5	23.08	45.81	-1.91			
10	72.31	48.04	4.46			
20	66.92	53.35	59.24			
IC_{50}	7.5	5	18			
(µg/ml)						
		В				
	Inhibition ratio % extract of Lentinula edodes					
CONC.		LC202				
µg/ml	MCF7 Hep-G2 HCT116					
	NICF /	nep-G2	nciilo			
2.5	32.05	67.88	33.76			
5	55.90	46.93	30.89			
10	59.49	2.51	32.48			
20	60.77	66.48	6.05			
IC_{50}	4.5	5	No cytotoxic			
(µg/ml)			effect			
		С				
	Inhibition ratio	% extract of <i>F. ve</i>	lutipes Fv5			
CONC.						
µg/ml	MCF7	Hep-G2	HCT116			
2.5	31.28	22.07	24.52			
5	20.51	41.06	17.20			
10	26.41	48.88	17.20			
20	29.49	55.03	52.23			
IC ₅₀	No cytotoxic effect	8	10			
1050						

		D		
	Inhibition ratio % extract of <i>F. velutipes Fv6</i>			
CONC. μg/ml	MCF7	Hep-G2	HCT116	
2.5	37.95	33.80	16.24	
5	45.64	55.03	4.46	
10	29.23	9.78	6.05	
20	53.33	67.60	29.62	
IC ₅₀ (µg/ml)	5	4	No cytotoxic effect	

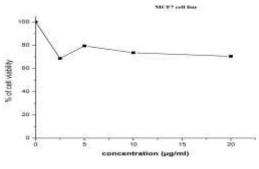
E						
CONC.	Inhibition ratio % extract of <i>F</i> . <i>velutipe13s</i>					
μg/ml	MCF7	Hep-G2	HCT116			
2.5	2.56	51.40	-33.76			
5	-16.15	46.09	-62.42			
10	3.59	17.88	-81.53			
20	-8.72	66.20	-81.53			
IC ₅₀	No cytotoxic	7	No cytotoxic			
(µg/ml	activity		activity			
)						



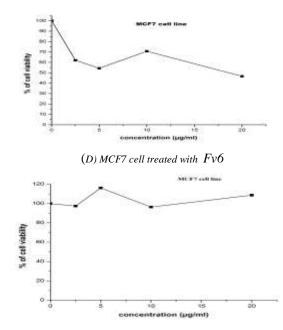
(A) MCF7 cell treated with LC2141



(B) MCF7 cell treated with LC202h

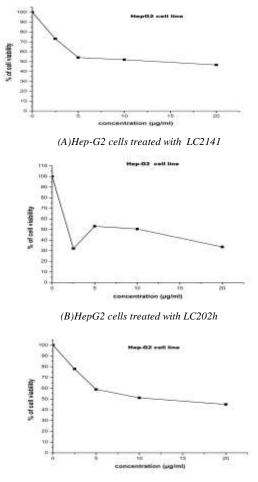


(C) MCF7 cell treated with Fv5

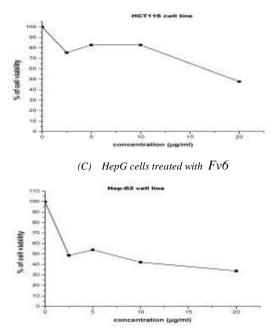


(E) MCF7 cell treated with Fv13

Fig. 1. (A-E) Inhibition ratios of MCF7 cell lines with IPS extract of the five studied Mashrom strain using concentration range from $2.5 - 20 \mu g/ml$



(C)HepG2 cells treated with Fv5



(E) HepG2 cells treated with Fv13

Fig. 2. (A-E) Inhibition ratios of HepG2 cell lines with IPS extract of the five studied Mashrom strain using concentration range from 2.5 - 20µg/ml

Only hot water extract of L. edodes LC2141 and F. velutipes 5 revealed strong inhibitions against HCT116 colon cell line with inhibition ratios of 59.24 % and 52.23% respectively at concentration of $20\mu g/ml$ (table 1). This result was consistent with previous investigations which confirmed that the polysaccharides or their complexes could have cytotoxicity or cytostatic effects on various tumor cell lines in vitro whereas they were less toxic on normal cells [23]-[25]. Also, our results was similar to reports observed by Ahmed et al., [26] that water extracts of F. velutipes fruiting bodies showed highest inhibition effect against three tumor cells (HepG-2, HCT 116 and HeLa) with IC50 ranging between 5-16 $\mu g/ml$

Our results demonstrated that mushroom extracts can be a good source for antitumor substances, and these results are consistent with reports indicating that mushrooms extracts had antitumor effects. For examples, low-molecular weight mushroom substances have been studied, and reported to interact with particular intracellular signaling pathways related to processes such as inflammation, cell differentiation and survival, apoptosis, angiogenesis, tumor progression and metastasis [27]. Many polysaccharide or polysaccharide protein complexes isolated from mushrooms exhibit antitumor activity [28]. Lentinan, a polysaccharide isolated from water extract of L. edodes has strong antitumor activities against stomach cancer (29, 30). Also, Finimundy et al. [31] reported high inhibitory activities against both HeLa and Hep G2 cell lines by water extracts of L. edodes

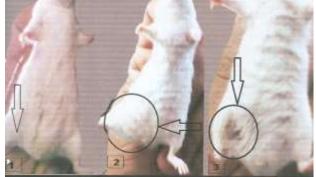
The difference in the antitumor activity may be attributed to their different molecular weights, monosaccharide composition, and charge characteristics. The results suggested that F. velutipes 6 polysaccharide extract had potential for investigation for antitumor activities. So, it was selected for in vivo studies

Tumor Inhibition Effects

The anti-tumor activity of a polysaccharide is usually believed to be a consequence of the stimulation of the cell-mediated immune response [32]. Mice transplanted Ehrlich carcinoma cells was used to evaluate the effects of F. velutipes 6 polysaccharide extract in vivo

The antitumor activity of F. velutipes 6 polysaccharide extract against Ehrlich carcinoma tumor in the mice model was significant. As shown in fig. 3, the mean tumor volume reached 0.942 mm3 in the saline-treated control 6 weeks after tumor implantation. In contrast, a significant inhibition of tumor growth was observed in tested mice upon oral administration of 200 mg/kg F. velutipes 6 polysaccharide extract for 10 days before tumor implantation with the inhibition ratio of 99.34% (mean tumor volume was 0.006mm3), while 4 out of 15 mice showed complete tumor regression

Group with oral injection after tumor implantation of F. velutipes 6 polysaccharide extract inhibited tumor growth was even more effective than group injected with extract before tumor implantation. The inhibition ratio of 99.79 % (mean tumor volume was 0.002mm3), while 5 out of 15 mice showed complete tumor regression in this group



Effects of polysaccharide extract from F.velutipes Fv19 on 1-Negative control group, 2- Positive control group, & 3- Mice administrated polysaccharide extract after tumor induction

Effect of the F. velutipes 6 polysaccharide on Body Weight in Mice

Before embarking on the experiments, the body weights of all the groups were 20.0 ± 2.0 g. There was no significant difference in body weight of each group (p > 0.05). During the experiments, the appetite and activity of each animal in these groups were better than the mice with solid tumor only. The results showed little change in final body weight in group injected with extract before tumor implantation while significant increase (p > 0.05) in final weight in group injected with extract after tumor implantation. Especially in the solid tumor group, many mice exhibited ruffled fur, lethargy and loss of appetite, which suggested some toxic effect of on mice. On the other hand, the body weights in the group injected with extract only showed no significant change (p < 0.05) in body weight as compared to those of the normal control group with increase in activity and survival of mice. The results showed that there was no significant toxic effect of the F. velutipes 6 polysaccharide to Ehrlich implanted mice. Also, water extract of F. velutipes 6 increased the survival rate of the mice- bearing

tumor and their activity in comparison with positive control

IV. CONCLUSION

This study demonstrated that hot water extract of the mycelia from five different strains of mushroom, revealed cytotoxic activity against different cancer cell lines. Cytotoxicity assay suggest the polysaccharide of *F. velutipes 6* exhibited strong antitumor activities so, it was chosen for *in vivo* experiment. Characterization of most active hot water extract and the immunomodulating activity of *F. velutipes 6* polysaccharide will be further investigated in future work.

REFERENCES

- Kidd, P.M. The use of mushroom glucans and proteoglycans in cancer treatment. Altern Med Rev 5:4–27 (2000).
- [2] Zaidman, B.Z., Yassin, M., Mahajna, J. and Wasser, S.P. Medicinal mushroom modulators of molecular targets as cancer therapeutics. ApplMicrobiolBiotechnol67:453–468 (2005).
- [3] Wasser, S.P. Medicinal mushrooms as a source of antitumor and immunomodulating polysaccharides. Appl Microbiol Biotechnol 60:258–274 (2002). https://doi.org/10.1007/s00253-002-1076-7
- [4] Moradali, M.F., Mostafavi, H., Ghods, S. and Hedjaroude, G.A. immunomodulating and anticancer agents in the realm of macromycetes fungi (macrofungi). Int Immunopharmacol 7:701–724 (2007).
- https://doi.org/10.1016/j.intimp.2007.01.008
 [5] Zhang, M., Cui, S.W., Cheung, P.C.K. and Wang, Q. Antitumor polysaccharides from mushrooms: a review on their isolation process, structural characteristics and antitumor activity. Trends Food SciTechnol18:4–19 (2007).
 https://doi.org/10.1016/j.tifs.2006.07.013
- [6] Ferreira, I.C.F.R., Vaz, J.A., Vasconcelos, M.H. and Martins, A. Compounds from wild mushrooms with antitumor potential. Anti-Cancer Agents Med Chem10:424–436 (2010). https://doi.org/10.2174/1871520611009050424
- [7] Peng, Y., Zhang, L., Zeng, F. and Xu, Y. Structure and antitumor activity of extracellular polysaccharides from mycelium. Carbohydr Polym 54:297–303 (2003).

https://doi.org/10.1016/S0144-8617(03)00190-5

- [8] Caili, F., Haijun, T., Tongyi, C., Yi, L. and Quan hong, L. Some properties of an acidic protein-bound polysaccharide from the fruit of pumpkin. Food Chem 100:944–947 (2007). https://doi.org/10.1016/j.foodchem.2005.10.049
- [9] Lull, C., Wichers, H.J. and Savelkoul, H.F.J. Anti inflammatory and immunomodulating properties of fungal metabolites. Mediat Inflamm 2:63–80 (2005).

https://doi.org/10.1155/MI.2005.63

- [10] 1Cui, J., Goh, K.K.T., Archer, R. and Singh, H. Characterisation and bioactivity of protein-bound polysaccharides from submerged culture fermentation of Coriolus versicolor Wr-74 and ATCC-20545 strains. J Ind Microbiol Biotechnol 34:393–402 (2007). https://doi.org/10.1007/s10295-007-0209-5
- [11] Sun, Y. and Liu, J. Purification, structure and immunobiological activity of a water-soluble polysaccharide from the fruiting body of Pleurotus ostreatus. Bioresour Technol 100:983–986 (2009). https://doi.org/10.1016/j.biortech.2008.09.036
- [12] Cui, F. J., Li, Y., Xu, Z. H., Xu, H. Y., Sun, K., and Tao, W. Y.. Optimization of the medium composition for production of mycelial biomass and exo -polymer by Grifola frondosa GF9801 using response surface methodology. Bioresource Technology, 97, 1209–1216 (2006). https://doi.org/10.1016/j.biortech.2005.05.005
- [13] Wang, J. C., Hu, S. H., Liang, Z. C., and Yeh, C. J.. Optimization for the production of water-soluble polysaccharide from Pleurotus citrinopileatus in submerged culture and its antitumor effect. Applied Microbiology and Biotechnology, 67:759–766(2005). https://doi.org/10.1007/s00253-004-1833-x
- [14] Hu, S. H., Liang, Z. C., Chia, Y. C., Lien, J. L., Chen, K. S., and Lee, M. Y., et al. Anti hyperlipidemic and antioxidant effects of extracts from Pleurotu scitrinopileatus. Journal of Agricultural and Food Chemistry, 54: 2103–2110 (2006).

https://doi.org/10.1021/jf052890d

- [15] -Duan, Y.Q., Xing, Z.C. and Xu, J.W. Screening of a high yield polysaccharide strain from ten edible and medicinal fungi and optimization of its culture conditions. Res. J. Biotech., 8:11-15(2013).
- [16] Wu, X.; Xu, R.; Ren, Q.; Bai, J. and Zhao, J.: Factors affecting extracellular and intracellular polysaccharide production in submerged cultivation of Tricholoma Mongolicum. Afr. J. Microbiol. Res. 6(5): 909-916(2012).
- [17] Mizuno, T. The extraction and development of antitumor-active polysaccharides from medicinal mushrooms in Japan. International Journal of Medicinal Mushrooms 1: 9-29 (1999). https://doi.org/10.1615/IntJMedMushrooms.v1.i1.20
- [18] Cui, J.; Guo, S. and Xiao P.: Antitumor and antimicrobial activities of endophytic fungi from medicinal parts of Aquilaria sinensis. J Zhejiang Univ Sci B. 12:385–392 (2011). https://doi.org/10.1631/jzus.B1000330
- [19] Rajendran, L.; Rajagopal, K.; Subbarayan, K.; Ulagappan, K.; Sampath, A. and Karthik, G.: Efficiency of fungal taxol on human liver carcinoma cell lines. American J Res Communication. 1:112-121(2013).
- [20] Evidente, A.; Kornienko, A.; Cimmino, A.; Andolfi, A.; Lefranc, F.; Mathieud, V. and Kiss, R.: Fungal metabolites with anticancer activity. Nat Prod Rep. 31:617-627 (2014). https://doi.org/10.1039/c3np70078j
- [21] Wang, L.W.; Zhang, Y.L.; Lin, F.C.; Hu, Y.Z. and Zhang, C.L.: Natural products with antitumor activity from endophytic fungi. Mini Rev Med Chem.11:1056-74 (2011). https://doi.org/10.2174/138955711797247716
- [22] Cui, F.J., Tao, W.Y., Xu, Z.H., Guo, W.J., Xu, H.Y., Ao, Z.H., Jin, J. and Wei, Y.Q. Structural analysis of antitumor heteropolysaccharide GFPS1b from the cultured mycelia of Grifola frondosa GF9801. Bioresource Technology 98, 395–401 (2007). https://doi.org/10.1016/j.biortech.2005.12.015
- [23] Cui, F.J., Li, Y., Xu, Y.Y. and Liu, Z.Q. et al., Induction of apoptosis in SGC-7901 cells by polysaccharide-peptide GFPS1b from the cultured mycelia of Grifola frondosa GF9801. Toxicology in Vitro 21 (2007) 417–427.

https://doi.org/10.1016/j.tiv.2006.10.004

- [24] Yang, X. Sit, W.H. Chan, D.K. and Wan, J.M. The cell death process of the anticancer agent polysaccharide-peptide (PSP) in human promyelocytic leukemic HL-60 cells .Oncology Reports 13: 1201–1210(2005). https://doi.org/10.3892/or.13.6.1201
- [25] Daba, A.S. and Ezeronye, O.U. Anti-cancer effect of polysaccharides isolated from higher basidiomycetes mushrooms. African Journal of Biotechnology 2 (12) 2672–678 (2003).
- [26] Ahmed, Y.; Jennifer, S.; Fang-Sheng, W.; Hussien, E.; Fathy, H. and Mahmoud, E.: Effectiveness of different solvents extracts from edible mushrooms in inhibiting the growth of tumor cells. Cancer Biology, 4(4) (2014).
- [27] Petrova, R.D.: New scientific approaches to cancer treatment: can medicinal mushrooms defeat the curse of the century? Int J Med Mushrooms. 14:1-20 (2012).

https://doi.org/10.1615/IntJMedMushr.v14.i1.10

- [28] Ma, L.; Chen, H.; Dong, P. and Lu, X.: Anti-inflammatory and anticancer activities of extracts and compounds from the mushroom Inonotus obliquus. Food Chem. 139:503- 508 (2013). https://doi.org/10.1016/j.foodchem.2013.01.030
- [29] Kim, H.Y.; Kim, J.H.; Yang, S.B.; Hong, S.G.; Lee, S.A.; Hwang, S.J.; Shin, K.S.; Suh, H.J. and Park, M.H.: A polysaccharide extracted from rice bran fermented with Lentinus edodes enhances natural killer cell activity and exhibits anticancer effects. J Med Food 10:25–31(2007). https://doi.org/10.1089/jmf.2006.116
- [30] Ina, K.; Kataoka, T. and Ando, T.: The use of lentinan for treating gastric cancer. Anticancer Agents Med Chem. 13:681-688 (2013). https://doi.org/10.2174/1871520611313050002
- [31] Finimundy, T.C.; Gambato, G.; Fontana, R.; Camassola, M.; Salvador, M.; Moura, S.; Hess, J.; Henriques; J.A.; Dillon, A.J. and Roesch-Ely, M.: Aqueous extracts of Lentinula edodes and Pleurotus sajor-caju exhibit high antioxidant capability and promising in vitro antitumor activity. Nutr Res. 33:76-84 (2013).

https://doi.org/10.1016/j.nutres.2012.11.005

[32] 32-Ooi, V. E., & Liu, F.: Immunomodulation and anti-cancer activity of polysaccharide–protein complexes. Current Medicinal Chemistry, 7, 715–729 (2000). https://doi.org/10.2174/0929867003374705



First A. Author and the other authors may include biographies at the end of regular papers. This author became a Member (M) of URUAR The first paragraph may contain a place and/or date of birth (list place, then date). Next, the author's educational background is listed. The degrees should be listed with type of degree in what field, which institution, city, state or country, and year degree was earned. The author's major field of study should be lower-cased.

The second paragraph uses the pronoun of the person (he or she) and not the author's last name. It lists military and work experience, including summer and fellowship jobs. Job titles are capitalized. The current job must have a location; previous positions may be listed without one. Information concerning previous publications may be included. Try not to list more than three books or published articles. The format for listing publishers of a book within the biography is: title of book (city, state: publisher name, year) similar to a reference. Current and previous research interests ends the paragraph.

The third paragraph begins with the author's title and last name (e.g., Dr. Smith, Prof. Jones, Mr. Kajor, Ms. Hunter). Finally, list any awards and work for committees and publications. If a photograph is provided, the biography will be indented around it. The photograph is placed at the top left of the biography. Personal hobbies will be deleted from the biography.